

Development of  
a novel culture-independent method for  
comprehensive detection and identification  
of beer-spoilage microorganisms

Masaki Shimokawa, Kazumaru Iijima, Koji Suzuki,  
Yasuo Motoyama, Hiromi Yamagishi.

Asahi Breweries, Ltd.

# Contents

1. Background

2. Purpose and strategy

~screening a membrane × pressure cycling technology~

3. Results

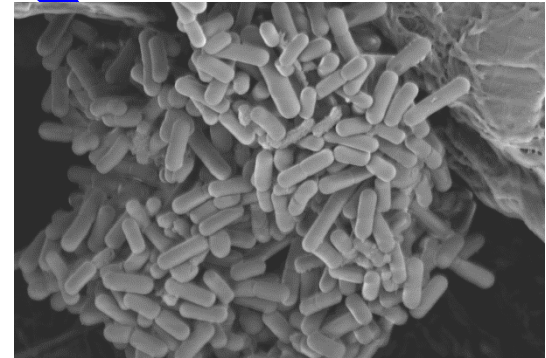
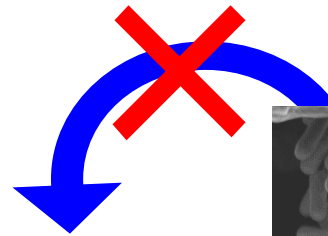
4. Discussion

5. Conclusion

# Features of Japanese beer

Unpasteurized beer has dominated the Japanese beer market.

Beer is a microbiologically stable beverage.



Beer-spoilage microorganisms

Stringent microbial quality control required.

# Beer-spoilage microorganisms

<i>Lactobacillus</i>	<i>Pediococcus</i>	<i>Pectinatus</i>
<i>L. brevis</i>	<i>Ped. damnosus</i>	<i>P. frisingensis</i>
<i>L. lindneri</i>	<i>Ped. claussenii</i>	<i>P. crevisiiphilus</i>
<i>L. paracollinoides</i>	<i>Ped. inopinatus</i>	<i>P. haikarae</i>
<i>L. backi</i>	other <i>Pediococcus</i>	Other <i>Pectinatus</i>
<i>L. coryniformis</i>	Wild yeasts	<i>Megasphaera</i>
<i>L. paucivorans</i>	<i>S. cerevisiae</i>	<i>M. cerevisiae</i>
<i>L. casei /paracasei</i>	<i>D. anomalla</i>	<i>M. paucivorans</i>
<i>L. plantarum</i>	<i>D. buruxellensis</i>	<i>M. sueciensis</i>
other <i>Lactobacilli</i>	<i>B. custersianus</i>	

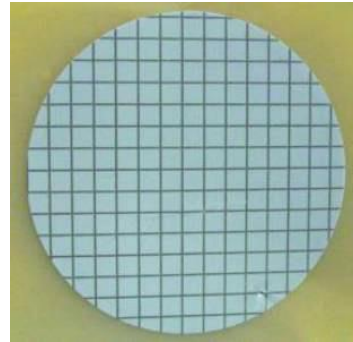
Currently new species are also emerging!

# Conventional microbiological QC test

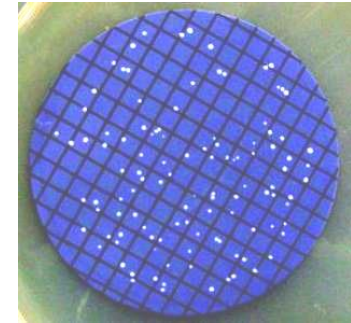
**Beer filtration**



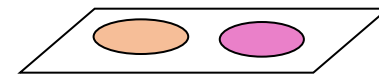
**Culture**



**Identification**



- Microscopic observations
- Gram staining
- Catalase activities
- PCR tests e.t.c...



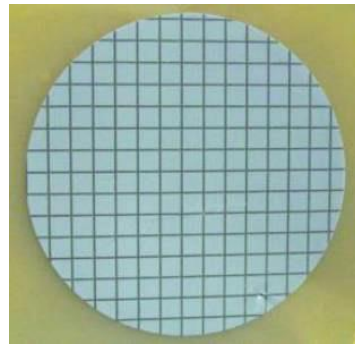
**Rate-limited process**

In some urgent cases, more rapid methods may be needed!

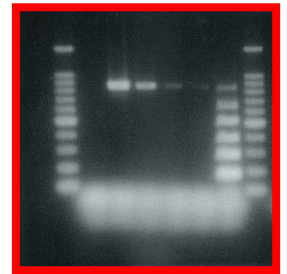
Would it be possible to detect without culture?



**Beer filtration**



**Extract DNA directly**



# Limitations of culture-independent method

	species	membrane	filtration volume	detection limit
Tsuchiya, Y. (1992)	<i>Lactobacillus brevis</i>	PVDF	250	30 cells/250 mL
Tsuchiya, Y. (1993)	<i>L.brevis</i>	Polycarbonate	250	1-9 cells/250 mL
DiMichele, L. J. (1993)	<i>Lactobacillus</i>	Polycarbonate	50	20 cells/mL
Satokari, R. (1997)	<i>Pectinatus</i>	Polycarbonate	100	20 cells/mL
Yasui, T. (1997)	<i>L.lindneri</i>	PVDF	100	63 cfu/100 mL

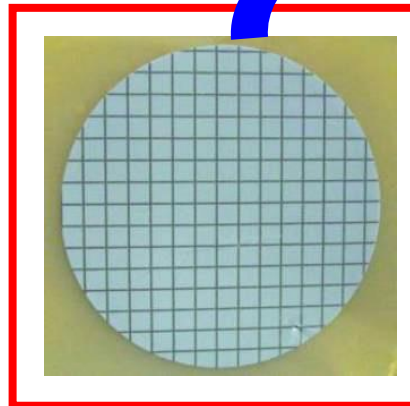
A comprehensive direct method with higher detection limits has been pursued for the past 20 years.

# Purpose and strategy of this study

## Purpose

Development of a comprehensive highly sensitive culture-independent detection method

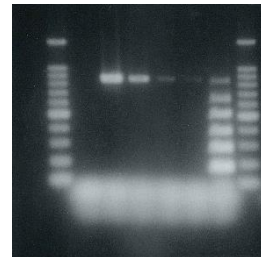
### Beer filtration



Increase  
filtration vol.



Extract DNA  
efficiently



Multiplex  
PCR

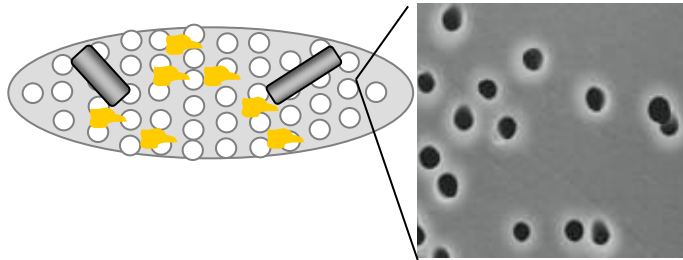


# Screening of a membrane

pros and cons of each membrane type

# Polycarbonate membrane

Polycarbonate  
(isopore type)



## Advantages

Sensitivity is relatively high.  
(approximately 10cells/membrane)

## Disadvantages

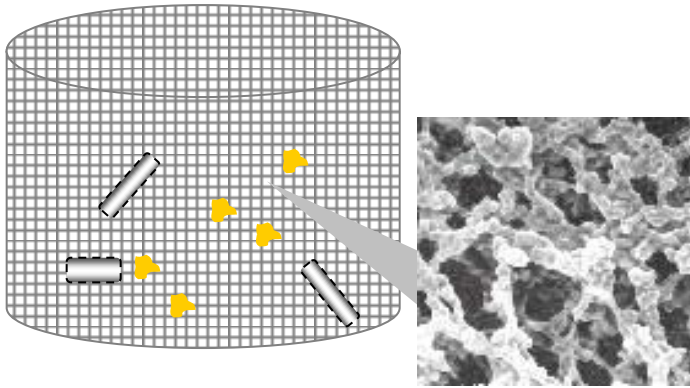
Filtration volume is limited.  
(up to 250ml)

Lotus root-like structure  
(**Bacteria trapped on the surface.**)

A trace level of beer spoilers cannot be detected

# Cellulose membrane

## Mixed cellulose ester



## Advantages

Filtration volume is larger.  
(3000ml is possible)

## Disadvantages

Sensitivity is low.  
(Approx. 100 cells/membrane)

## Mesh-like structure

(Bacteria trapped somewhere in the middle.)

Cellulose  
membrane



Cells are buried in membrane.

Cells and DNA must be recovered  
more efficiently.

Mixed cellulose ester membrane

×

Pressure cycling technology

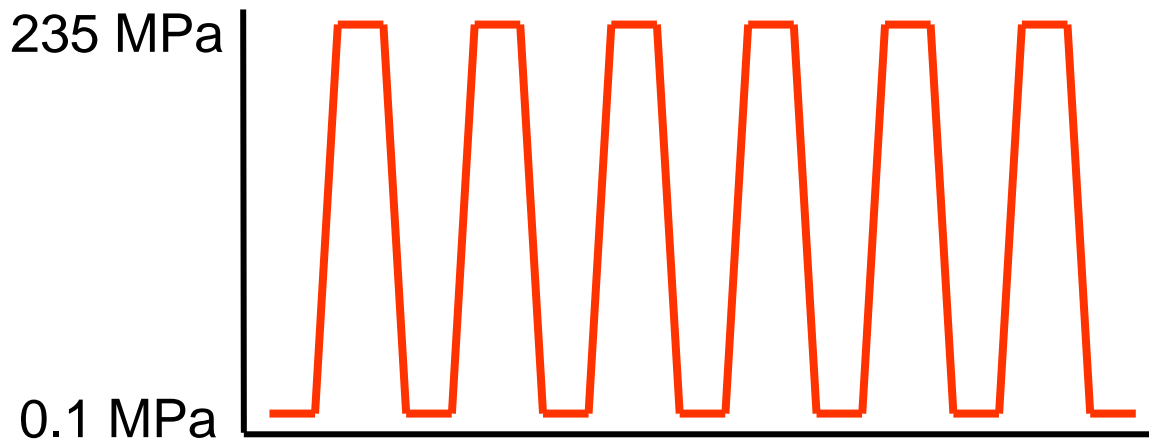
# Pressure cycling technology (PCT)

Barocyler™ NEP2320



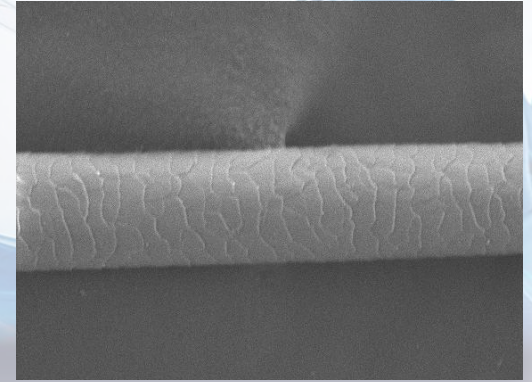
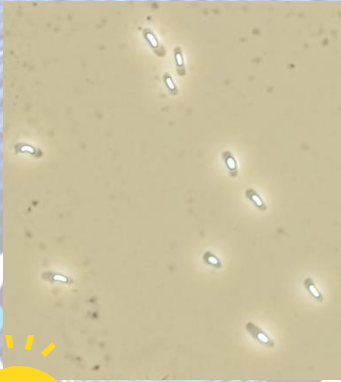
A novel approach for sample preparation method using alternating levels of hydrostatic pressure.

0.1 MPa → 235 MPa : less than 3sec  
235 MPa → 0.1 MPa : less than 1sec



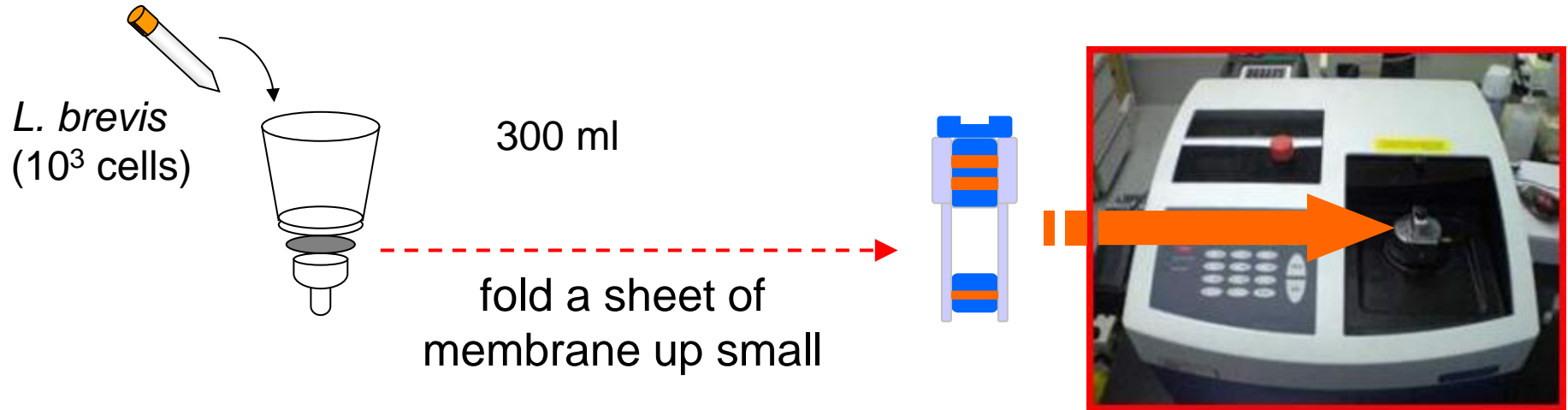
# Performance of Pressure Cycling Technology

Significant improvements in DNA yield from challenging biological and forensic samples using Pressure Cycling Technology.



**It might be possible to extract DNA from cells buried in membrane without disruption!**

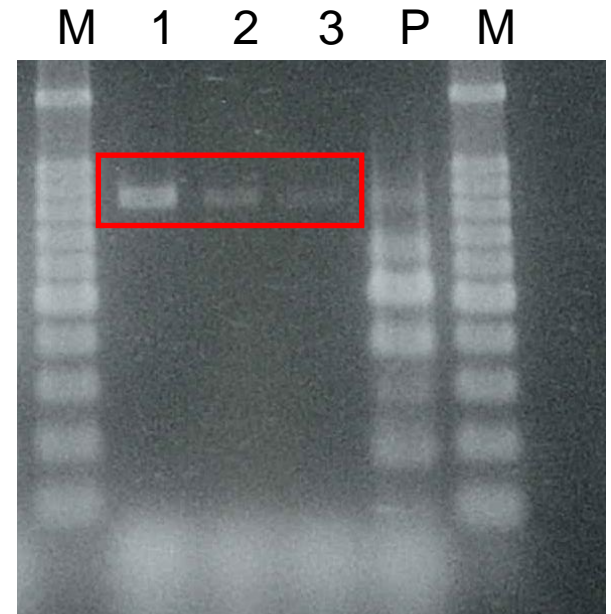
# The performance of PCT for DNA extraction



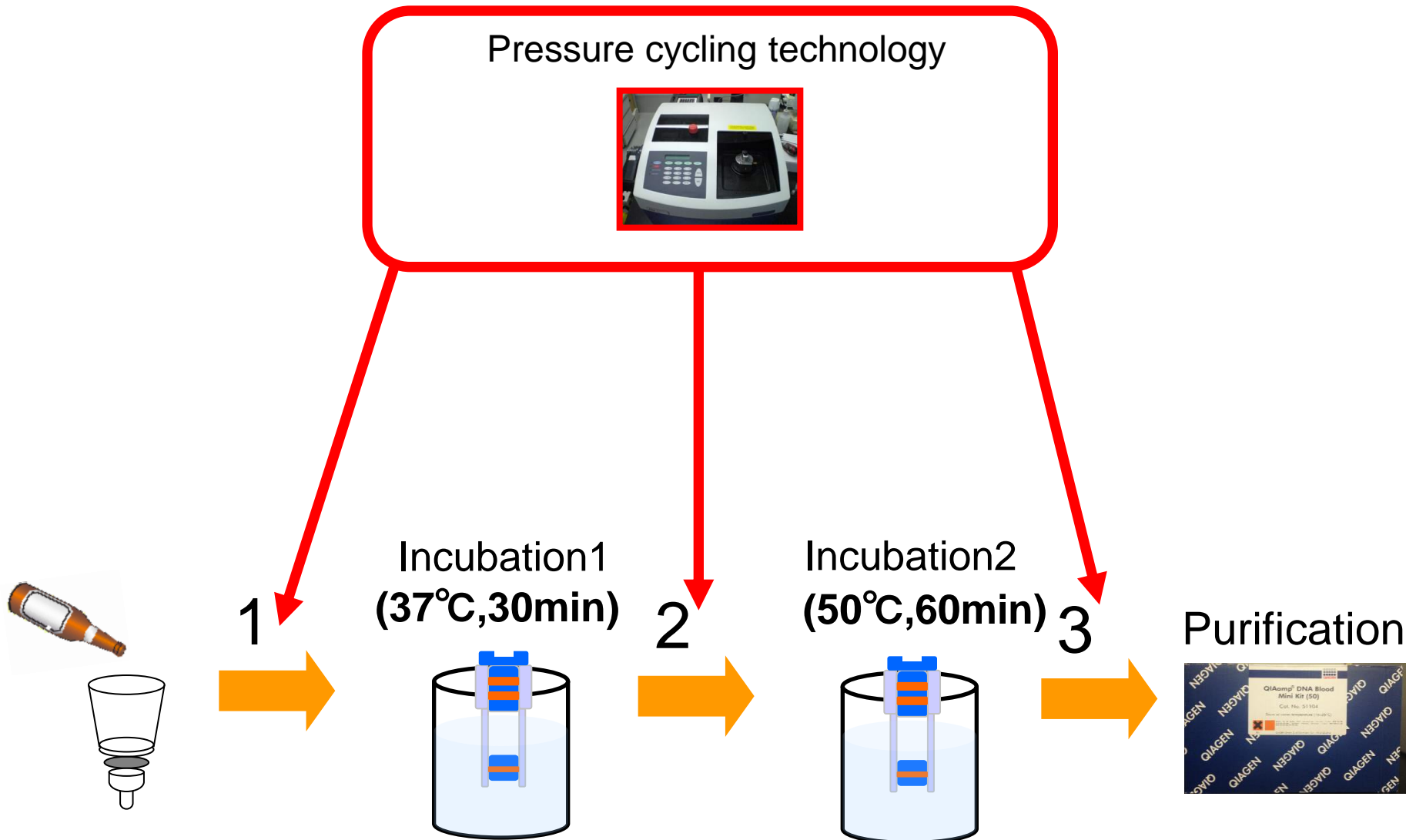
Three extraction methods were compared

1. PCT with enzymatic treatment
2. Only enzymatic treatment
3. Only PCT

PCT was used once in this protocol.

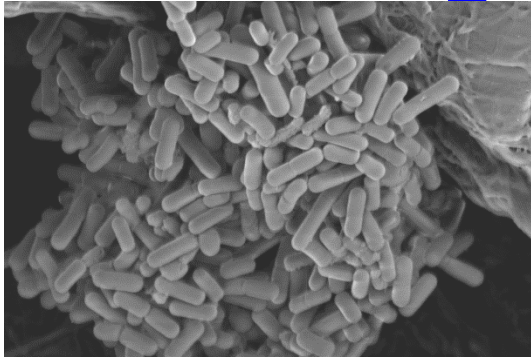


# Schematic flow of DNA extraction





# Evaluation of sensitivity



Beer spoilage microorganisms

*L. brevis*

*L. lindneri*

*L. paracollinoides*

Wild yeasts

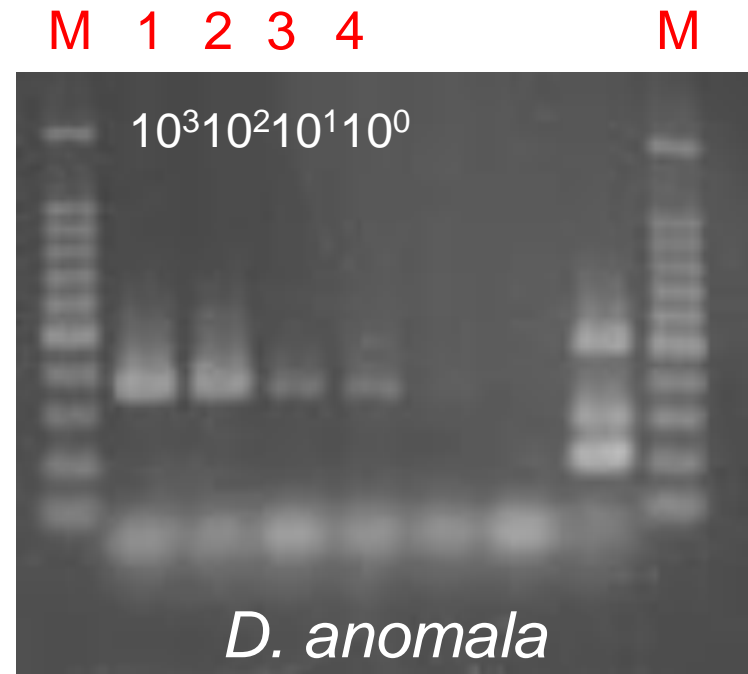
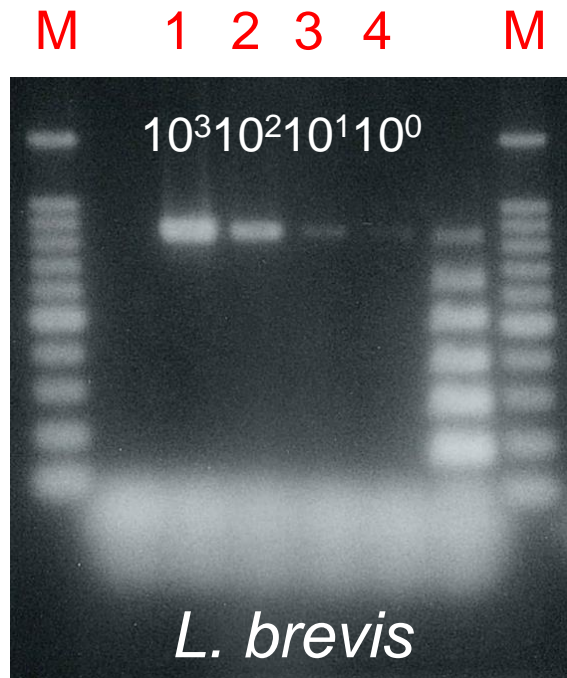


300 mL

**DNA extraction with pressure cycling technology**

# Evaluation of sensitivity for beer spoiler

The results of *L. brevis* and *D. anomala* are shown here as an example.



As for three *Lactobacillus* species and four wild yeast species, the sensitivity was shown to lie between  $10^0$  and  $10^1/300$  ml of beer.

# Mini summary 1

- The pressure cycling technology has enabled effective DNA extraction from the cells trapped within a cellulose membrane filter matrix.
- The detection limits of major beer-spoilage lactic acid bacteria, *L. brevis*, *L. lindneri*, *L. paracollinoides*, wild yeast (the genera *Saccharomyces* and *Dekkera/Brettanomyces*) species, were found to be as low as  $10^0$  cells/membrane (300ml beer).

Other lactic acid bacteria and strictly anaerobic bacteria are also reported as beer spoilers...

<i>Lactobacillus</i>	<i>Pediococcus</i>	<i>Pectinatus</i>
<i>L. brevis</i>	<i>Ped. damnosus</i>	<i>P. frisingensis</i>
<i>L. lindneri</i>	<i>Ped. claussenii</i>	<i>P. crevisiophilus</i>
<i>L. paracollinoides</i>	<i>Ped. inopinitus</i>	<i>P. haikarae</i>
<i>L. backi</i>	other <i>Pediococcus</i>	Other <i>Pectinatus</i>
<i>L. coryniformis</i>	<b>Wild yeasts</b>	<b>Megasphaera</b>
<i>L. paucivorans</i>	<i>S. cerevisiae</i>	<i>M. cerevisiae</i>
<i>L. rosei/parvacei</i>	<i>D. anomalla</i>	<i>M. paucivorans</i>
<i>L. plantarum</i>	<i>D. buruxellensis</i>	<i>M. custersiana</i>
other lactobacilli	<i>B. custersianus</i>	

**Pectinatus**  
10<sup>1-2</sup>

10<sup>1</sup>

10<sup>1</sup>

10<sup>1</sup>

cells/membrane

cells/membrane

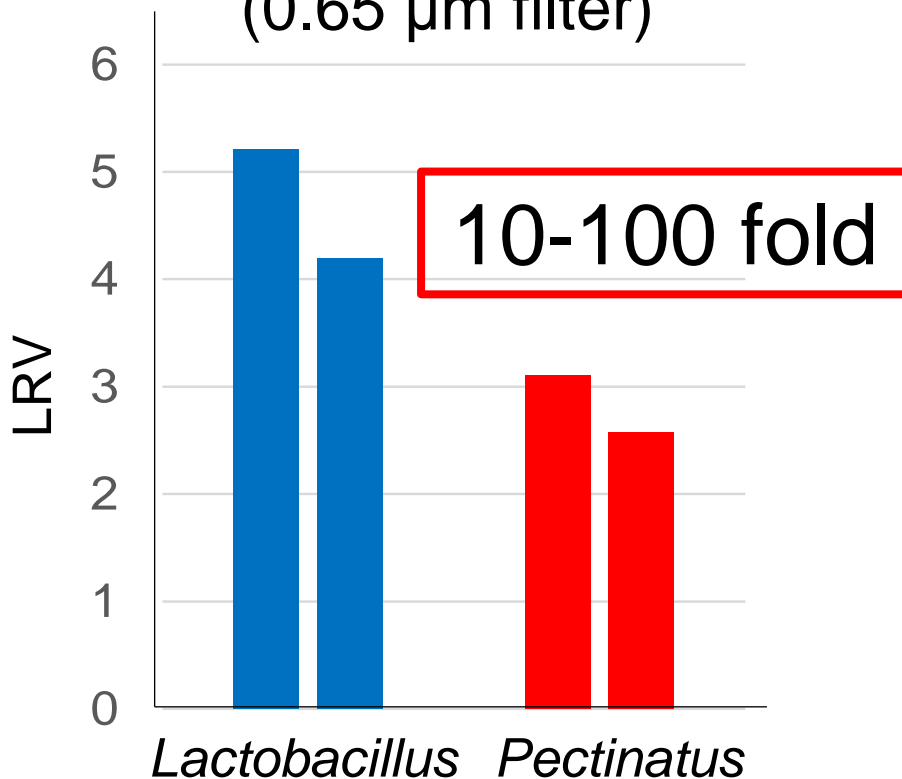
cells/membrane

cells/membrane

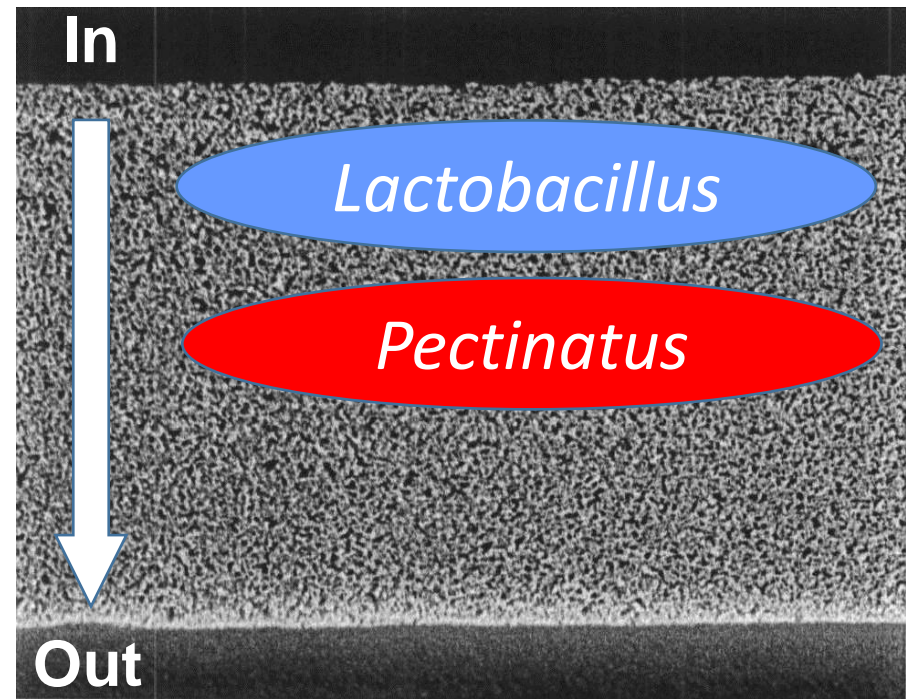
# Differences between *Pectinatus* and *Lactobacillus*

*Pectinatus* is reported to be more likely to pass through membrane filter than *Lactobacillus*.

Log Reduction Value; LRV  
(0.65  $\mu\text{m}$  filter)

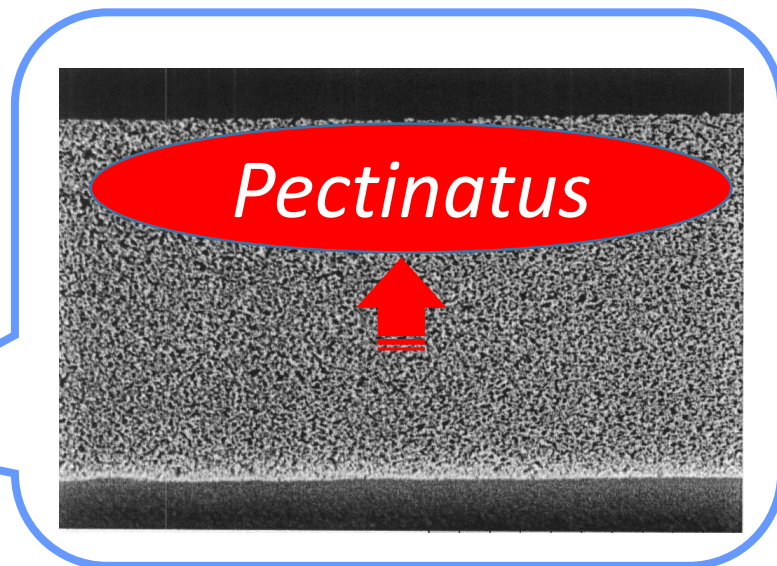
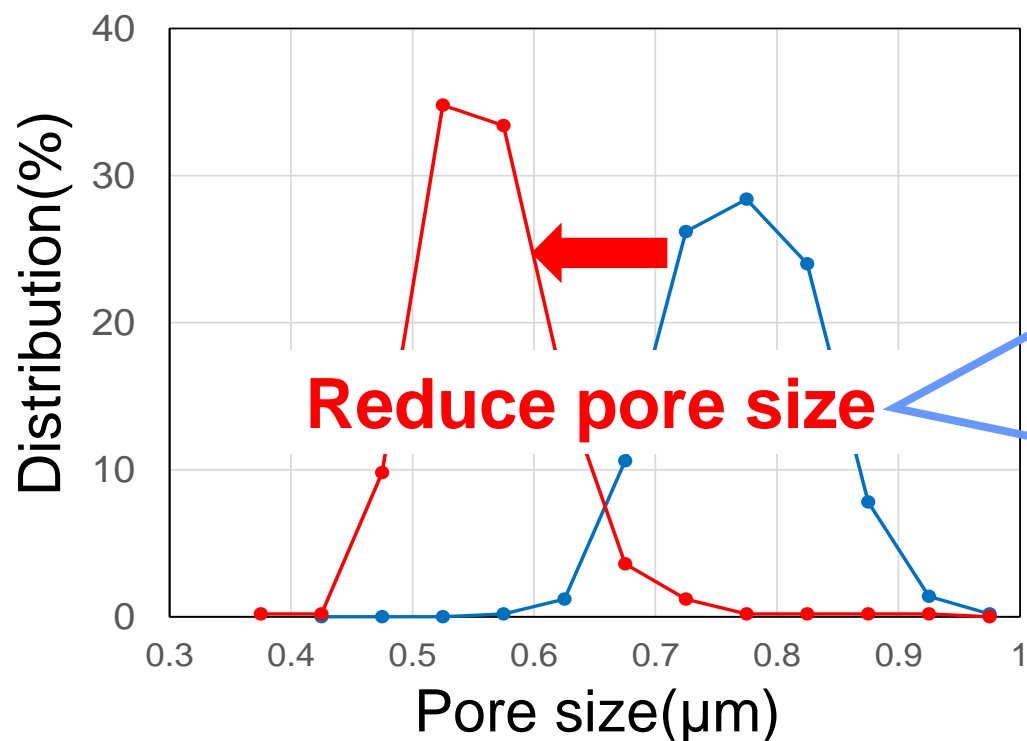


Cross-section structure  
of membrane



# Screening of a more optimal membrane

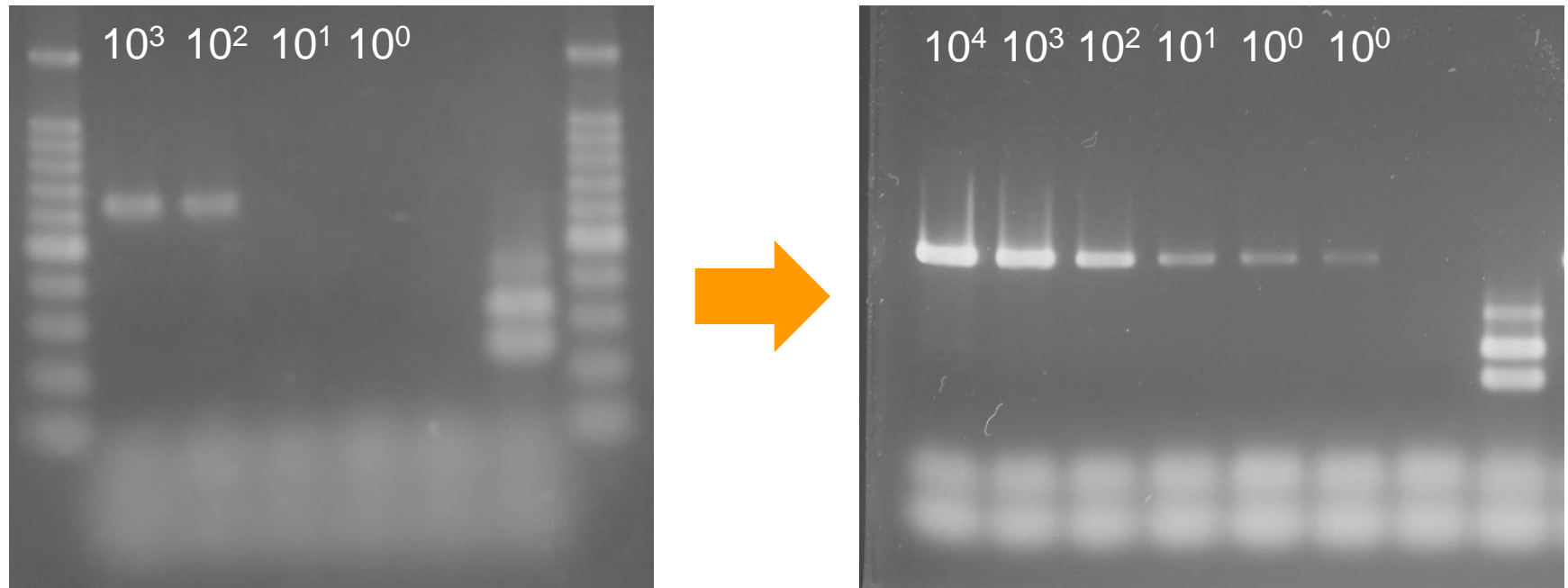
Optimization of pore size for cellulose membrane



Improvement of the DNA extraction efficiency  
by trapping the cells closer to the surface

# The result of optimization of membrane

The results of *P. cerevisiiphilus* are shown here as an example.

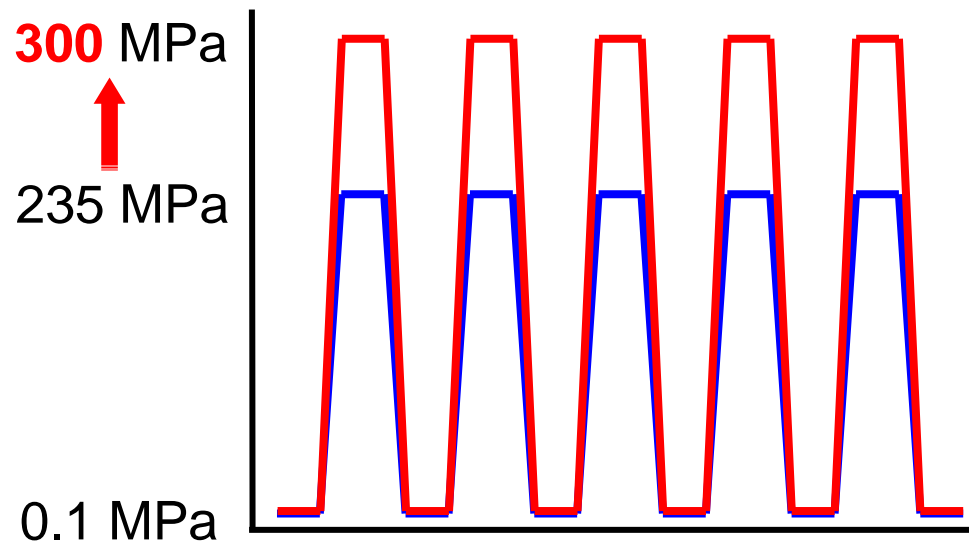
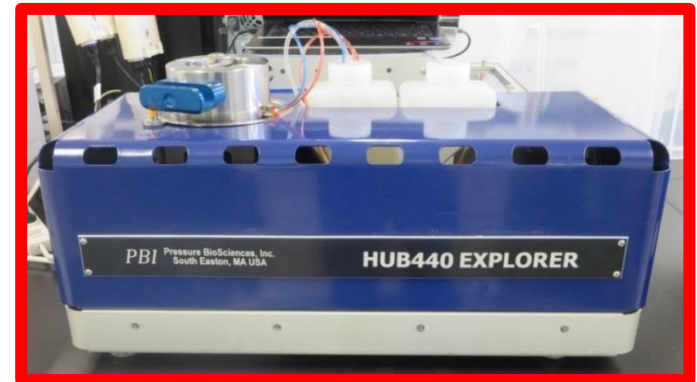


The modified approach was found to be applicable to all of the beer-spoilage *Pectinatus* species.



# To further improve extraction efficiency

Higher pressure (300 Mpa) model was adopted



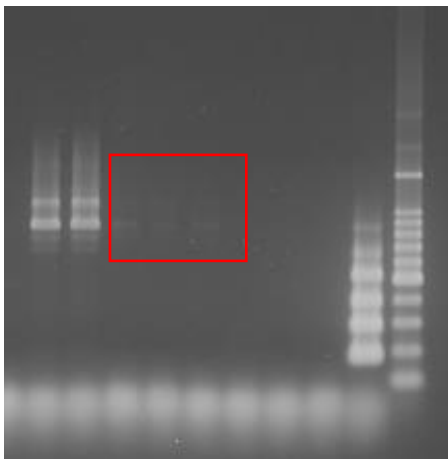


# To further improve sensitivity

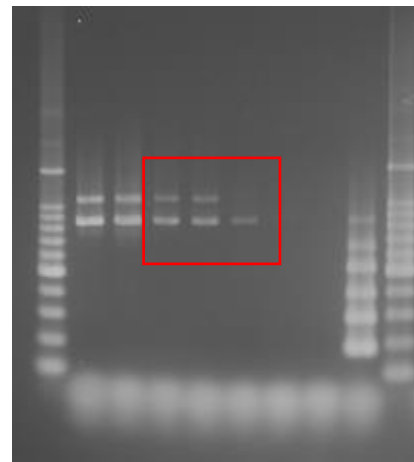
## Vacuum concentrator



Improves the sensitivity by evaporating DNA solution with vacuum concentrator.

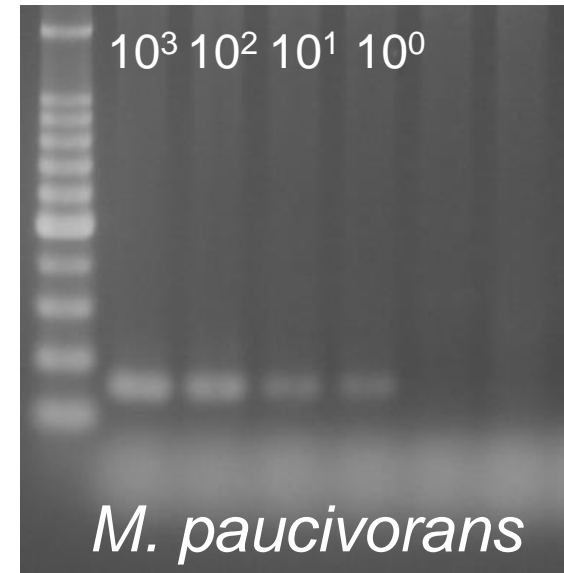
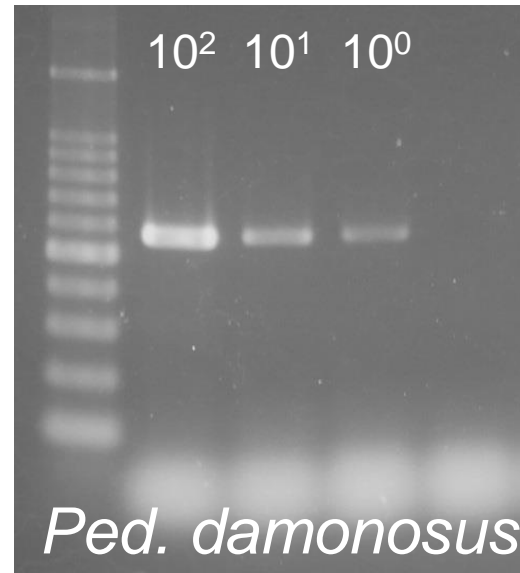
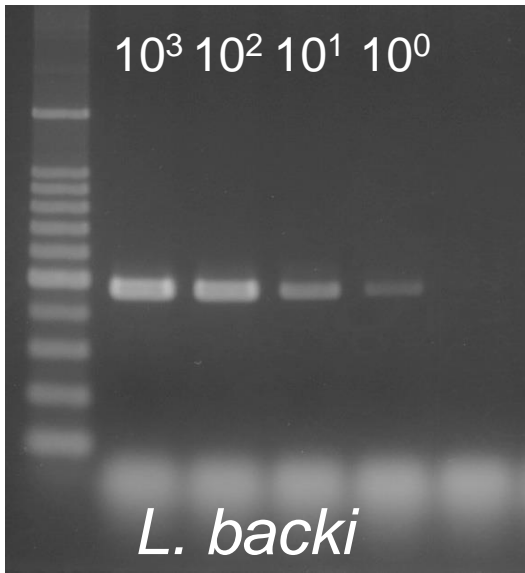


10-fold concentrated



Recovers a trace amount of DNA

# Result of using a wider range of beer spoilers



The identical detection limits were accomplished.

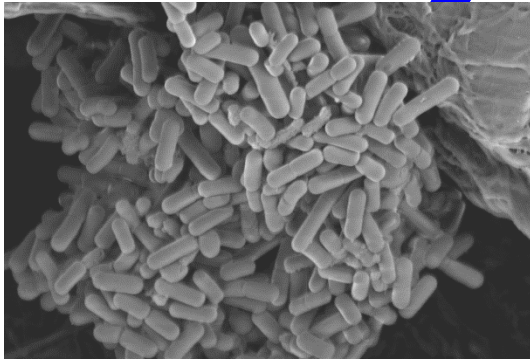
Our modified method was also shown to be applicable to other *Lactobacillus*, *Pediococcus* and *Megasphaera*.

# Mini summary 2

Series of measures in combination finally allow the detection of 22 species of beer-spoilage microorganisms with the detection limits of  $10^0$  cells/membrane

<b><i>Lactobacillus</i></b>	<b><i>Pediococcus</i></b>	<b><i>Pectinatus</i></b>
<i>L. brevis</i>	<i>Ped. damnosus</i>	<i>P. frisingensis</i>
<i>L. lindneri</i>	<i>Ped. claussenii</i>	<i>P. crevisiiphilus</i>
<i>L. paracollinoides</i>	<i>Ped. inopinatus</i>	<i>P. haikarae</i>
<i>L. backi</i>	<b>Wild yeasts</b>	<b><i>Megasphaera</i></b>
<i>L. coryniformis</i>	<i>S. cerevisiae</i>	<i>M. cerevisiae</i>
<i>L. paucivorans</i>	<i>D. anomalla</i>	<i>M. paucivorans</i>
<i>L. casei /paracasei</i>	<i>D. buruxellensis</i>	<i>M. sueciensis</i>
<i>L. plantarum</i>	<i>B. custersianus</i>	

**If filtration volume is increased up to 3000 mL**



*L.brevis*

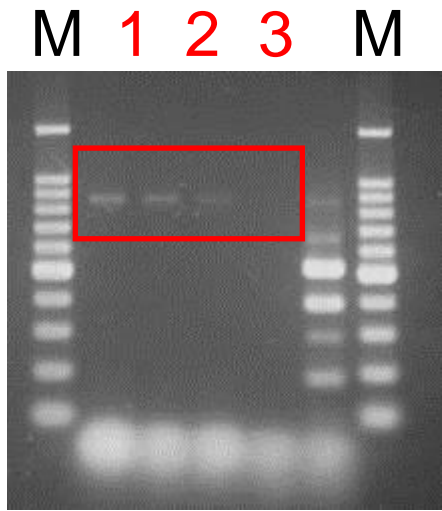


3000 mL



**DNA extraction with pressure cycling technology**

# Increased filtration volume



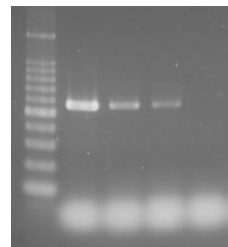
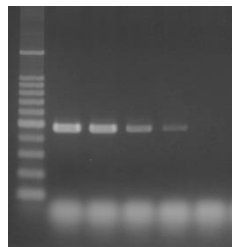
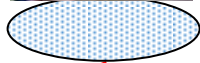
Inoculation level (cfu)

1	$10^2$ cells
2	$10^1$ cells
3	$10^0$ cells

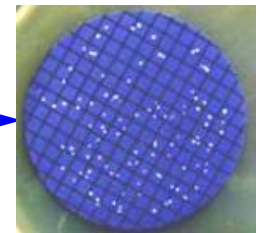


Detection limit  
 $10^0$  cell/3,000ml

A trace amount of bacteria in larger volume of beer was detected.



Direct detection  
(8hrs)



Culture

# Summary



- The pressure cycling technology has enabled effective DNA extraction from cells trapped within a cellulose membrane filter matrix.
- Series of measures in combination finally allow the detection of 22 species of beer-spoilage microorganisms with detection limits of  $10^0$  cells/membrane.
- Our method is able to cope with an extremely low level of contamination ( $10^0$  cell/3000ml-beer).



**Comprehensive detection and identification of low levels of beer-spoilage microorganisms is achieved by direct PCR**

Thank you for your attention

